

## Research Article

# Vaccine against MUC1 Antigen Expressed in Inflammatory Bowel Disease and Cancer Lessens Colonic Inflammation and Prevents Progression to Colitis-Associated Colon Cancer

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## Abstract

Association of chronic inflammation with an increased risk of cancer is well established, but the contributions of innate versus adaptive immunity are not fully delineated. There has furthermore been little consideration of the role played by chronic inflammation-associated antigens, including cancer antigens, and the possibility of using them as vaccines to lower the cancer risk. We studied the human tumor antigen MUC1 which is abnormally expressed in colon cancers and also in inflammatory bowel disease (IBD) that gives rise to colitis-associated colon cancer (CACC). Using our new mouse model of MUC1<sup>+</sup> IBD that progresses to CACC, interleukin-10 knockout mice crossed with MUC1 transgenic mice, we show that vaccination against MUC1 delays IBD and prevents progression to CACC. One mechanism is the induction of MUC1-specific adaptive immunity (anti-MUC1 IgG and anti-MUC1 CTL), which seems to eliminate abnormal MUC1<sup>+</sup> cells in IBD colons. The other mechanism is the change in the local and the systemic microenvironments. Compared with IBD in vaccinated mice, IBD in control mice is dominated by larger numbers of neutrophils in the colon and myeloid-derived suppressor cells in the spleen, which can compromise adaptive immunity and facilitate tumor growth. This suggests that the tumor-promoting microenvironment of chronic inflammation can be converted to a tumor-inhibiting environment by increasing adaptive immunity against a disease-associated antigen. *Cancer Prev Res*; 3(4): 438–46. ©2010 AACR.

## Introduction

Patients with chronic inflammatory disorders have an increased incidence of cancer development at the affected site (1). There is evidence that chronic inflammation may serve as the carcinogenic event (1–3). On the other hand, oncogenic events, such as *RET/PTC* gene rearrangement found in thyroid cancer (4) or activation of *Myc* and *Ras* found in many cancers, can initiate inflammatory transcriptional programs in transformed cells or the surrounding stromal cells, generating chronic inflammation at the tumor site (5, 6). Thus, regardless of the order of events, inflammation preceding cancer or cancer preceding inflammation, the result can be a tumor-promoting chronic inflammatory microenvironment. Chronically inflamed sites contain a diverse population of leukocytes that

express and secrete an assortment of mediators that foster cell proliferation and genomic instability (2, 7). We wanted to know whether this microenvironment could be altered by the presence of strong adaptive immunity specific for an antigen expressed at the disease site. Our hypothesis was that early in inflammation, effective priming of the adaptive immunity to many potential neoantigens at the affected site, or a prompt recall response to some of those antigens encountered previously, may clear the site and prevent chronic inflammation altogether, or prevent development of cancers that would be expected to express some of the same neoantigens. We have tested this hypothesis in a mouse model of inflammatory bowel disease (IBD) and colitis-associated colon cancer (CACC) that we previously described (8). As a target antigen for the adaptive immune response, we used the mucin glycoprotein MUC1 (9).

MUC1 is expressed at low levels on the apical surface of ductal epithelial cells in several organs including on the colonic epithelium (10). The extracellular domain is dominated by a variable number of tandem repeat (VNTR) region composed of an average of 80 to 200 repeats that are 20 amino acids in length. The 20-amino acid peptide sequence HGVTSAPDTRPAPGSTAPPA has five potential O-glycosylation sites that on healthy epithelia are heavily glycosylated. In the majority of adenocarcinomas and their precursor lesions, and in various inflammatory diseases

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including IBD, MUC1 is overexpressed and its VNTR region is profoundly hypoglycosylated (8, 9, 11–14). The reduced glycosylation exposes the peptide backbone, resulting in peptide epitopes as well as truncated novel glycopeptide epitopes that are processed and presented to the immune system (15–17). Furthermore, we and others discovered that this hypoglycosylated MUC1 has chemotactic properties for the cells of the innate immune system that have multiple receptors capable of binding this abnormal form (16, 18–20). Abnormal expression of MUC1 in IBD thus provides a source of predictable disease-associated neoepitopes that could exert important influence on the innate immune system as well as be targets of adaptive immunity.

We previously characterized a mouse model of MUC1-positive human IBD (MUC1<sup>+</sup> IBD; ref. 8) derived by crossing human MUC1 transgenic mice (21) that faithfully replicate tissue-specific MUC1 expression in humans, with interleukin-10 knockout (IL-10<sup>-/-</sup>) mice that develop spontaneous IBD. *De novo* expression of abnormally glycosylated MUC1 on affected colonocytes resulted in worse inflammation and in increased cancer development (8). In the current study, we used the same mouse model to show that vaccine-elicited adaptive immunity to MUC1 early in the disease process leads to delayed and tempered development of IBD, changes in the inflammatory microenvironment, and complete protection from colon cancer.

## Materials and Methods

### Mice

IL-10<sup>-/-</sup> mice on a C57BL/6 background were purchased from The Jackson Laboratory, and MUC1 transgenic mice on a C57BL/6 background were originally purchased from Dr. S.J. Gendler (The Mayo Clinic, Scottsdale, AZ) and are bred at the University of Pittsburgh. IL-10<sup>-/-</sup> mice (homozygous for the IL-10 deletion) develop spontaneous colitis with pathologic changes similar to human IBD. Human MUC1 transgenic mice express full-length human MUC1 in the same spatial and tissue distribution as the endogenous protein in humans (21). The MUC1<sup>+</sup>/IL-10<sup>-/-</sup> progeny (25% of total) generated by crossing IL-10<sup>-/-</sup> mice with MUC1 transgenic mice develops MUC1-positive IBD (8). Littermates that do not carry this genotype are used as controls, thus keeping the living conditions of all mouse groups equal. All experiments were approved by the Institutional Animal Care and Use Committee of the University of Pittsburgh (Pittsburgh, PA).

### PCR genotyping

PCR was used to identify the *MUC1* transgene as well as the presence or absence of the *IL-10* gene. The primer pairs for MUC1 transgenic mice were 5'-CTGCCAGCCATAG-GACCAAG-3' and 5'-CTCCACGTCGTGGACATTGATG-3'. The IL-10 primers were 5'-GTGGGTGCAGTTATTG-TCTTCCCG, 5'-GCCTTCAGTATAAAAGGGGGACC, and 5'-CCTGCGTGCAATCCATCTTG-3'. The PCR product of

each reaction was analyzed on a 1% agarose gel. MUC1 amplification resulted in a 500-bp fragment, and IL-10 amplification resulted in a 200-bp fragment if IL-10<sup>+/+</sup>, 200- and 450-bp fragments if IL-10<sup>+/-</sup>, and 450-bp fragment if IL-10<sup>-/-</sup>.

### Peptide synthesis and *in vitro* glycosylation

The TnMUC1 100-mer peptide used for immunization corresponds to five tandem repeats of a 20-amino acid sequence HGVTSAPDTRPAPGSTAPPA from the extracellular VNTR region of MUC1. Enzymatic addition of GalNAc to the peptide was done using recombinant UDP-GalNAc polypeptide *N*-acetyl-galactosaminyltransferases rGalNAc-T1 as previously described (22). The MUC1 peptide MUC1-9-5N9L and glycopeptide MUC1-10-5GalNAc correspond to the extracellular VNTR region of MUC1. The peptides were synthesized at the University of Pittsburgh Genomics and Proteomics Core Laboratories.

### Vaccine protocol

Mice in the vaccine group received 20  $\mu$ L intranasally (10  $\mu$ L/nare) of 30  $\mu$ g of TnMUC1 100-mer peptide mixed with 3  $\mu$ g of adjuvant E6020 in PBS. Adjuvant-treated mice received 3  $\mu$ g of adjuvant E6020 in PBS. Mice received vaccine or adjuvant between 5 and 6 wk of age and were boosted twice at 2-wk intervals. The adjuvant E6020 is a synthetic, attenuated Toll-like receptor-4 agonist provided by Eisai Research Institute, Andover, MA (23–25).

### Histology and inflammation scores

Colon was removed immediately after sacrifice, cut longitudinally, washed in cold 1 $\times$  PBS, fixed in 5% buffered formalin, and embedded in paraffin. Sections (5  $\mu$ m thick) were stained with H&E. Inflammation scores (0–4) were determined by a gastrointestinal pathologist who was blinded to the experimental protocol using criteria previously reported (8). Briefly, 20 to 40 separate microscopic fields were evaluated for each mouse and graded 0 to 4. The total inflammation score for each sample was determined by taking the sum of the fields divided by the number of fields.

### Immunohistochemistry

Tissue paraffin sections (5  $\mu$ m thick) were deparaffinized by baking overnight at 59°C. Endogenous peroxidase activity was eliminated by treatment with 30% H<sub>2</sub>O<sub>2</sub> for 15 min at room temperature. Antigen retrieval was done by microwave heating in 0.1% citrate buffer. Nonspecific binding sites were blocked with Protein Blocking Agent (Thermo-Shandon). The anti-MUC1 antibody HMPV (BD Pharmingen) recognizes all forms of MUC1 by binding the epitope APDTR in the VNTR region in a glycosylation-independent manner. The anti-MUC1 antibody VU-4H5 (Santa Cruz Biotechnology) recognizes the epitope APDTRPAP in the VNTR region of hypoglycosylated MUC1. Staining was done by the avidin-biotin-peroxidase method with a commercial kit (Vectastain

ABC kit, Vector Laboratories). Color development was done using a 3,3'-diaminobenzidine kit (BD Pharmingen).

### Microscopy and image acquisition

Histology sections were observed using an Olympus BX40 microscope. Images were acquired using a Leica DFC420 camera and Leica Application Suite version 2.7.1 R1.

### Lactoferrin ELISA

Fecal samples were measured for the concentration of lactoferrin as previously described (26). Briefly, fecal pellets between 15 and 30 mg were homogenized in 500  $\mu$ L of collection buffer. Collection buffer is 0.1% deoxycholate (Sigma) in PBS, 10  $\mu$ mol/L leupeptin (Sigma), 1.6  $\mu$ mol/L pepstatin A (Sigma), and 5  $\mu$ g/mL aprotinin (Sigma). Ninety-six-well Maxisorb plates (Fisher) were coated with 100  $\mu$ L/well of fecal homogenate or human lactoferrin standards (Sigma) and incubated overnight at 4°C. Plates were washed with PBS/0.05% Tween 20 and blocked for 1 h with bovine serum albumin. Rabbit anti-human lactoferrin antibody (Sigma) was diluted 1:500 in collection buffer and added at 100  $\mu$ L/well and incubated for 2 h at room temperature. Horseradish peroxidase-labeled goat anti-rabbit antibody (100  $\mu$ L/well) was added and incubated for 30 min at room temperature. For color development, 100  $\mu$ L/well of 2-nitro-5-thiobenzoate substrate (BD OptEIA) was added and incubated in the dark for 10 to 12 min. The reaction was stopped with 50  $\mu$ L/well of sulfuric acid. The plates were read at 450 nm. Fecal lactoferrin concentration is based on the human lactoferrin standard curve and dividing the lactoferrin (ng) by the weight of the starting fecal sample. This gives nanogram of lactoferrin per gram of feces.

### MUC1-specific ELISA

Blood was collected at 5 wk of age before treatment and at the time of sacrifice. Serum was tested for the presence of MUC1-specific antibodies using a MUC1-specific ELISA. Briefly, 96-well plates were coated overnight at 4°C with 10  $\mu$ g/mL of MUC1 100-mer peptide in PBS. Plates were washed with PBS/0.5% Tween 20 and incubated with mouse serum diluted at 1:50 for 1 h at room temperature. After three washes with 0.5% Tween 20, plates were incubated with goat anti-mouse peroxidase-conjugated secondary antibody (Sigma) for 1 h at room temperature. Plates were washed thrice with 0.5% Tween 20 and incubated with substrate reagent A&B (BD OptEIA, BD Biosciences) for 15 min. The reaction was stopped with 2.5 mol/L sulfuric acid, and absorbance was read at 450 nm.

### Chromium release assay

RMA and RMA-MUC1 target cells ( $10^6$ ) were labeled for 1 h with radioactive sodium chromate ( $\text{Na}_2\text{CrO}_4$ ), washed thrice with DMEM, resuspended at  $10^6$  cells per mL, and plated in 96-well V-bottomed microtiter plate at 2,000

per well in 100  $\mu$ L. Effector cells were added in triplicate in DMEM at  $10^5$  per well for 4 h. Supernatant was harvested and analyzed on a Cobra II auto-gamma counter (Perkin-Elmer). Specific lysis was calculated as (lysis – spontaneous lysis)/(total lysis – spontaneous lysis). Average of triplicates was plotted.

### Flow cytometry

Isolated cells from lymph nodes and spleens were washed and resuspended in fluorescence-activated cell sorting buffer (2% fetal bovine serum in PBS) and plated at  $0.5 \times 10^6$  to  $1 \times 10^6$  per well. Surface Fc receptors were blocked with the addition of anti-CD16 (BD Biosciences) and incubated for 20 min. Cells were stained with anti-CD3, anti-CD4, anti-CD8, anti-Gr1, and anti-CD11b (BD Biosciences). DimerX, Soluble Dimeric Mouse H-2Kb:Ig Fusion Protein (BD Biosciences) was used according to the manufacturer's protocol to detect MUC1-specific CD8 T cells. Cells were analyzed on a LSR II flow cytometer (BD Biosciences), running FACSDiva software.

### Statistical analysis

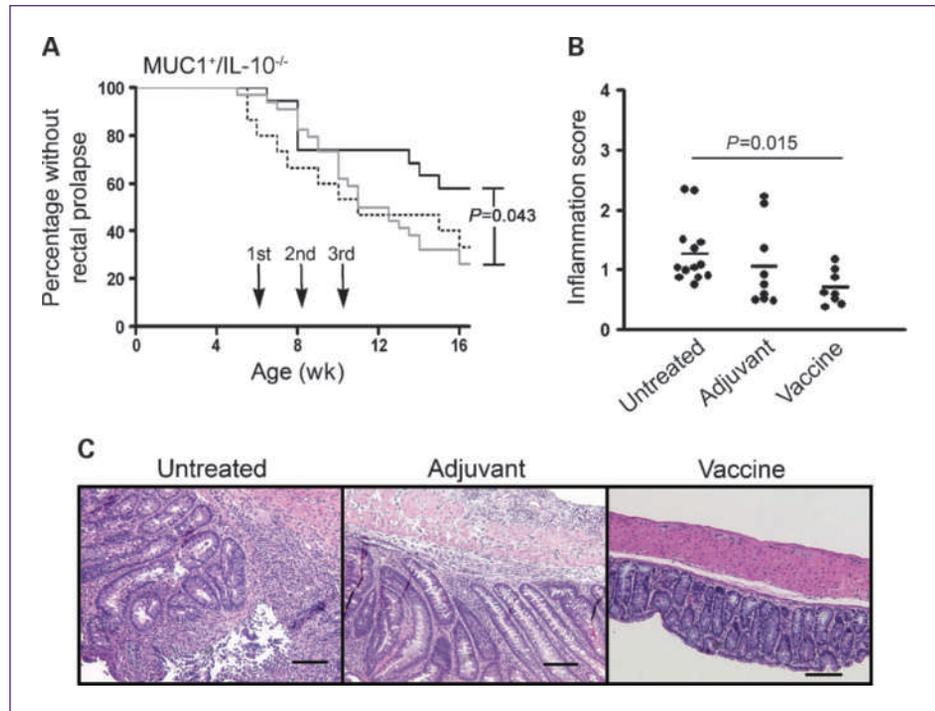
We conducted statistical analyses with GraphPad software. We analyzed Kaplan-Meier curves with the log-rank test. We calculated mean scores and statistical significance with Student's *t* test (unpaired two-tailed) and Fisher's exact test (two-tailed).

## Results

### MUC1 vaccine slows down IBD progression

MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice develop spontaneous IBD around 6 to 8 weeks of age. Unlike IBD that develops in IL-10<sup>-/-</sup> mice, which has been used as one of the major models of human IBD, the disease that arises in MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice overexpresses the abnormal, hypoglycosylated MUC1 characteristic of, and important for, development of human IBD (8). MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice were started on a course of vaccination at 5 to 6 weeks of age. The vaccine consisted of a synthetic 100-mer glycopeptide TnMUC1 corresponding to five 20-amino acid tandem repeats of MUC1, glycosylated *in vitro* to express the tumor-associated glycan GalNAc, plus E6020 adjuvant, a synthetic, attenuated Toll-like receptor-4 agonist that promotes both systemic and mucosal immunity (23–25). Mice received nasal administration of either the vaccine or the adjuvant alone as control and were boosted twice at 2-week intervals. We chose nasal administration to promote mucosal immunity capable of homing to the colonic mucosa. As an additional control, we included untreated mice. The three groups of mice were monitored until 16 weeks of age for the development of rectal prolapse as the first external symptom of IBD. Figure 1A shows that mice that received the vaccine developed rectal prolapse significantly later ( $P = 0.043$ ) compared with control mice. There was no significant difference between the two control groups. The difference between the vaccine group and the control groups emerged ~9 weeks of age, at which

**Fig. 1.** Vaccination of MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice with a MUC1 vaccine slows down IBD development and decreases intestinal inflammation. A, Kaplan-Meier curves representing development of rectal prolapse in vaccinated (solid line; *n* = 19), adjuvant-treated (dotted line; *n* = 16), or untreated (gray line; *n* = 34) mice. Arrows, administration of the vaccine or adjuvant alone at 6, 8, and 10 wk of age. *P* = 0.043, log-rank test. B, colonic inflammation scores for mice that received vaccine (*n* = 8), adjuvant alone (*n* = 9), or remained untreated (*n* = 13), sacrificed at 16 wk of age. *P* = 0.015, unpaired two-tailed Student's *t* test. C, representative H&E sections of colons from an untreated, an adjuvant-treated, and a vaccinated MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mouse. Scale bar, 50  $\mu$ m.



time cases of rectal prolapse in the vaccine-treated group reached a plateau, whereas mice in the control groups continued to develop rectal prolapse. This plateau started after the second administration of the vaccine.

We did the same experiment in IL-10<sup>-/-</sup> mice that do not express MUC1. As we previously published (8), in the absence of MUC1, the disease kinetics are different in these mice, but we could nevertheless use them as controls for the antigen (MUC1)-specific effects of the vaccine. These mice also received nasal administration of the vaccine or adjuvant alone, or remained untreated, and were followed up to 16 weeks of age for the development of rectal prolapse. There was no statistically significant difference in disease progression between the vaccinated and the control groups (data not shown).

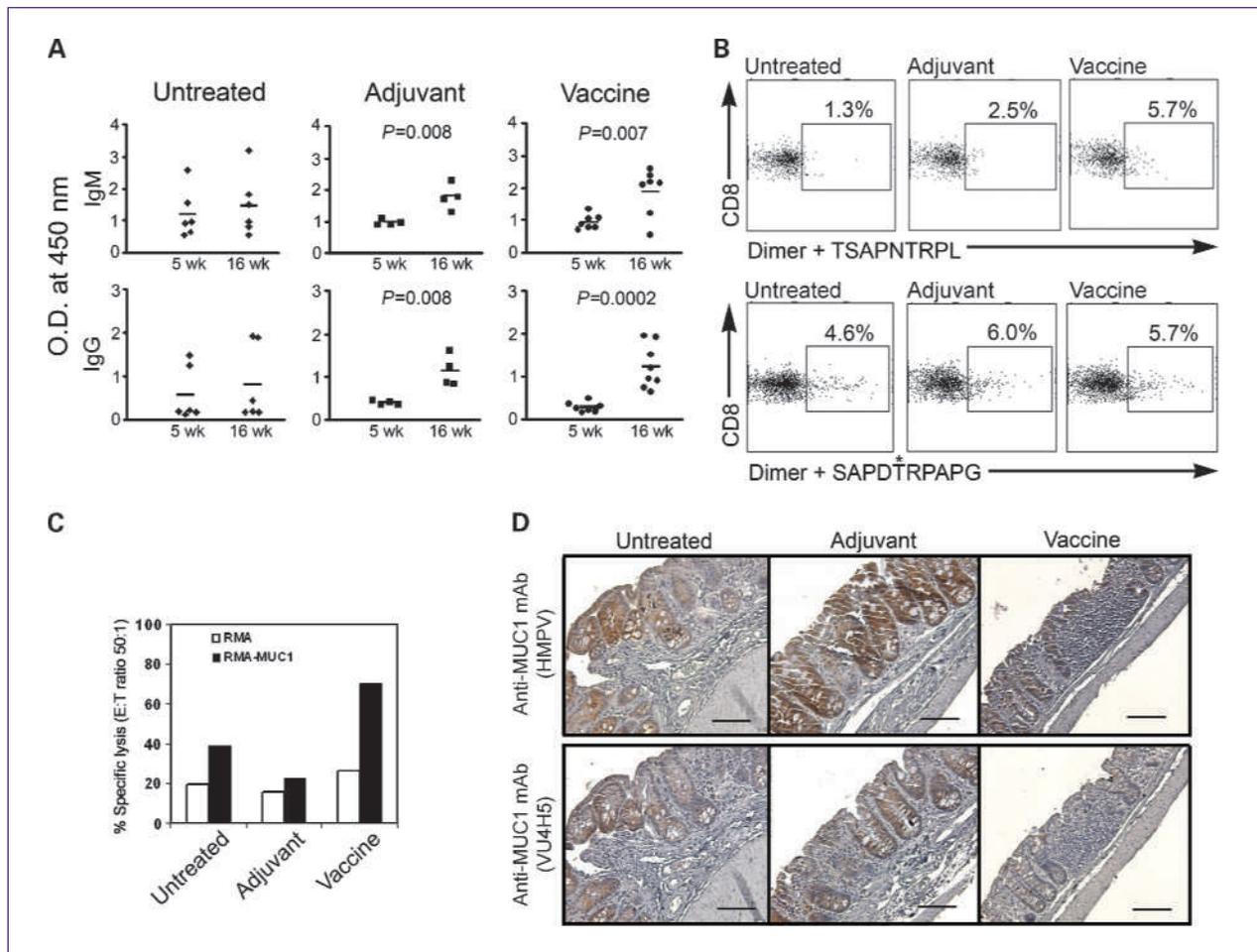
The differences between groups observed in MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice by monitoring the external signs of IBD were confirmed by pathology. Colons from each treatment group were collected at the time of sacrifice (16 weeks of age) and scored for inflammation grade by a pathologist blinded to the experimental protocol. Mice that received the vaccine had significantly lower inflammation scores (mean = 0.699) compared with untreated mice (mean = 1.256; *P* = 0.015; Fig. 1B). Although mice that received adjuvant alone also had higher inflammation scores (mean = 1.042) than the vaccinated mice, the difference did not reach statistical significance. This difference, however, was biologically significant. Representative tissue sections from a randomly selected mouse in each group (Fig. 1C), viewed at same magnification, show increased cellular infiltrate and pronounced thickening of the colon from an untreated and

an adjuvant-treated mouse compared with a vaccinated mouse.

#### Induction of MUC1-specific immunity in response to IBD versus the vaccine

We evaluated anti-MUC1 immunity in all three groups of mice that was either generated spontaneously in response to its abnormal expression in IBD, further enhanced in the presence of adjuvant alone, or elicited by the MUC1 vaccine. For measuring humoral responses, serum was collected at 5 to 6 weeks of age (before treatment) and again at 16 weeks of age, 2 weeks following the last injection in the treated groups. There was a significant increase in the ELISA absorbance values for anti-IgM in posttreatment samples compared with pretreatment serum levels for both the vaccinated mice (*P* = 0.007) and the adjuvant-treated mice (*P* = 0.008; Fig. 2A). We also detected a significant increase in the absorbance in posttreatment compared with pretreatment serum levels of anti-MUC1 IgG in adjuvant-treated mice (*P* = 0.008) and a highly significant increase in the vaccinated mice (*P* = 0.0002). Some of the untreated mice had high anti-MUC1 IgM and IgG responses at baseline, but in contrast to treated mice, there was no significant change between the two time points.

Using mouse MHC class I H2K<sup>b</sup> dimers (BD DimerX), MHC-Ig fusion proteins that detect antigen-specific T cells (27), we looked for the induction of MUC1-specific CD8 T cells. Dimers were loaded with either the 9-amino acid peptide MUC1-9-5N9L or the 10-amino acid glycopeptide MUC1-10-5GalNAc, both derived from the MUC1



**Fig. 2.** MUC1-specific immune responses in vaccinated versus control MUC1<sup>+/</sup>/IL-10<sup>-/-</sup> mice. **A**, MUC1-specific antibodies in serum of untreated ( $n = 6$ ), adjuvant-treated ( $n = 4$ ), and vaccinated ( $n = 7$ ) mice. Top, IgM isotype; bottom, IgG isotype. Absorbance (O.D.) was measured by ELISA at 450 nm. Serum samples were diluted 1:50.  $P$  value was determined by unpaired two-tailed Student's  $t$  test. **B**, splenocytes from untreated, adjuvant-treated, or vaccinated MUC1<sup>+/</sup>/IL-10<sup>-/-</sup> mice were stained with MUC1 peptide-loaded H2K<sup>b</sup>-Ig dimer. Representative dot plots from two independent experiments. **C**, results of standard <sup>51</sup>Cr release assay at an E:T ratio of 50:1. Effector cells are pooled lymph nodes from two mice per group. Targets are RMA (H2K<sup>b</sup>) or RMA-MUC1 (H2K<sup>b</sup>), transfected with human MUC1 cDNA. **D**, immunostaining (hematoxylin-counterstained) of colon sections with anti-MUC1 antibodies. Scale bar, 50  $\mu$ m.

VNTR region sequence HGVTSPADTRPAPGSTAPPA. In the peptide, the MHC class I anchor positions P5 and P9 in the native 9-amino acid peptide were changed from aspartic acid to asparagine and from alanine to leucine, respectively, converting TSAPDTRPA to TSAPNTRPL, to increase binding affinity for class I (28, 29). The MUC1 10-mer glycopeptide SAPDTRPAPG has the GalNAc glycan attached to the threonine at position 5. Dimers were loaded overnight with either peptide, mixed with pooled splenocytes from each group of mice, and analyzed by flow cytometry. Vaccinated mice had 5.7% of CD8 T cells that recognized the MUC1-9-5N9L peptide compared with 2.5% in the adjuvant group and 1.3% in the untreated group (Fig. 2B). Interestingly, all groups seemed to have CD8 T cells that recognized the MUC1-10-5GalNAc glycopeptide. We interpret this as an indication that MUC1 glycopeptide-specific CD8 T cells are generated in

response to the disease and that their induction is further promoted in the presence of adjuvant. The administration of the MUC1 vaccine, on the other hand, resulted in an increased frequency of CD8 T cells specific for both the peptide and the glycopeptide.

To test if these CD8 T cells were functional CTL, we again pooled splenocytes from vaccinated, adjuvant-treated, or untreated mice and used them as effector CTL against chromium-labeled targets, the mouse lymphoma cell line RMA, and the same cell line transfected with human MUC1 cDNA (RMA-MUC1). The highest lytic activity was found in spleens from mice that received the vaccine (Fig. 2C). This begged the question whether the presence of these CTLs in vaccinated animals may lead to elimination of cells expressing MUC1 from the colons of mice with IBD. MUC1 expression in colons from the vaccinated group and the controls was evaluated by immunohistochemistry

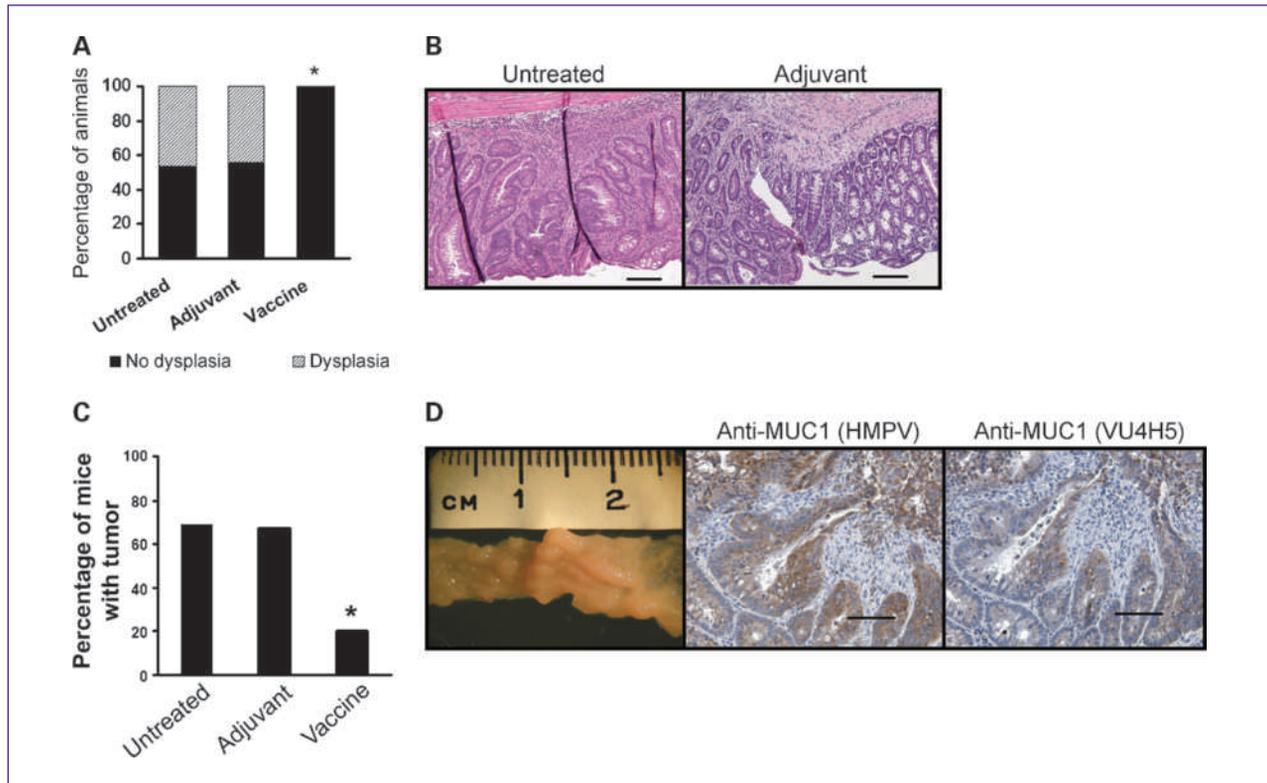
(Fig. 2D). Colon sections were stained with the anti-MUC1-specific monoclonal antibody HMPV that recognizes all forms of MUC1 (top) and with the anti-MUC1-specific monoclonal antibody VU4H5 that recognizes the abnormal hypoglycosylated form of MUC1 (bottom). Representative sections from each treatment group show that higher levels of MUC1 are observed in untreated and adjuvant-treated mouse colons compared with the vaccinated mouse (top). The MUC1 molecules in the untreated and adjuvant-treated mice are the hypoglycosylated forms. In contrast, very little if any hypoglycosylated MUC1 was detected in vaccinated mice (bottom).

### Vaccine-elicited MUC1 immunity prevents CACC

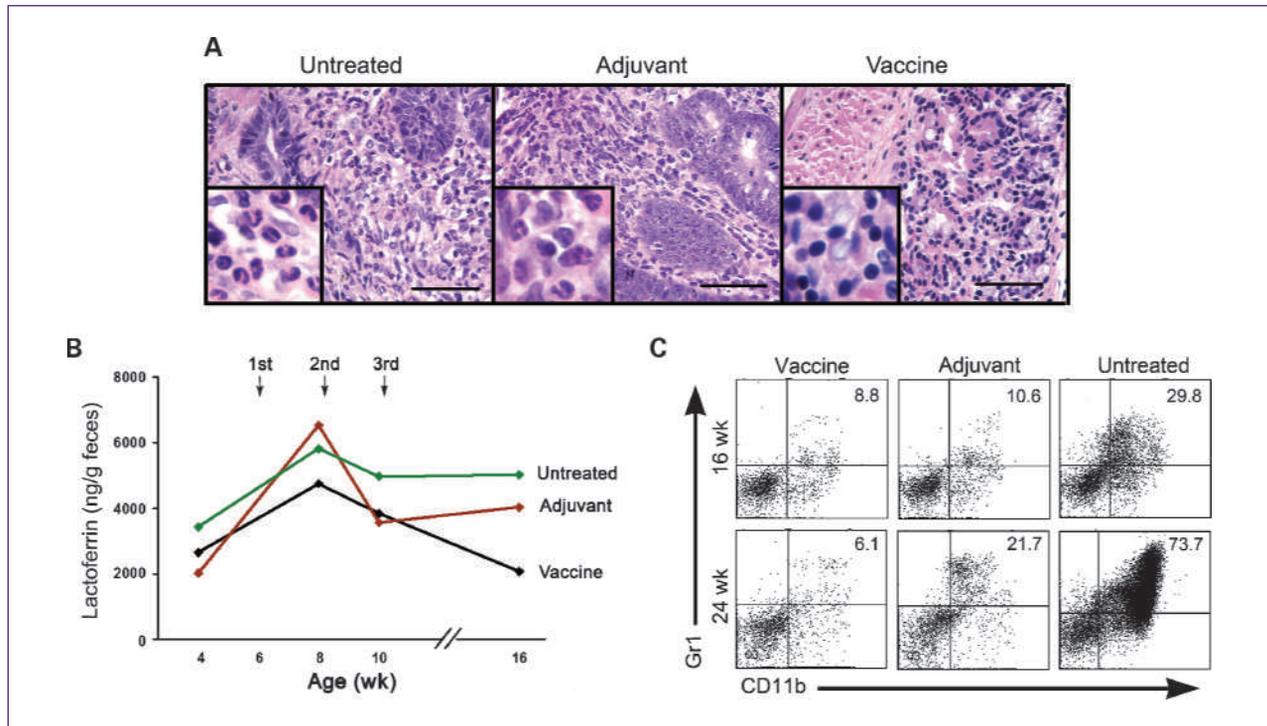
Given that our previous work showed that the majority of untreated MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice that developed IBD progressed to cancer (8), we evaluated the ability of the MUC1 vaccine to prevent early dysplastic lesions and CACC. The animals were matched by age and their colons were examined early in the course of disease for the development of colonic dysplasia. Alternatively, animals were grouped by the extent of their disease and the number of tumor foci evaluated in vaccinated mice relative to untreated mice or to animals that received adjuvant only.

At 16 weeks of age, there was no evidence of colonic dysplasia or neoplasia in eight of eight animals in the vaccinated group (Fig. 3A). In contrast, 6 of 13 (46%) untreated mice and 4 of 9 (44%) adjuvant-treated mice had colonic dysplasia. One mouse in the adjuvant group also had invasive colon cancer and one untreated mouse (not included in the graph) developed cancer at 12 weeks of age and had to be removed early from the protocol. Figure 3B shows examples of areas of dysplasia in colons of an untreated and adjuvant-treated mouse. We repeated the same analysis by focusing on colon tissue samples that had inflammation scores of >1.0. Of eight untreated mice in that category, six had dysplasia. The three adjuvant-treated mice all had dysplasia with one displaying neoplasia as well. Two mice in the vaccinated group had inflammation scores of >1.0, but neither of these colons had areas of dysplasia.

The second analysis was done on mice that had rectal prolapse for at least 2 weeks before sacrifice. Colons from such mice from all three groups were evaluated for the presence of neoplasia by a pathologist blinded to the experimental protocol. We found that vaccinated mice had statistically significantly fewer tumors compared with untreated mice ( $P = 0.041$ ; Fig. 3C). Eight of



**Fig. 3.** Vaccination against MUC1 prevents the development of dysplasia and CACC. A, percentage of untreated ( $n = 13$ ) and adjuvant-treated ( $n = 9$ ) mice with dysplasia at 16 wk of age. Vaccinated mice ( $n = 8$ ) had no areas of dysplasia. \*,  $P = 0.046$ , two-tailed Fisher's exact test. B, representative H&E-stained colon sections showing dysplastic lesions. Scale bar, 50  $\mu\text{m}$ . C, percentage of untreated ( $n = 16$ ), adjuvant-treated ( $n = 6$ ), or vaccinated ( $n = 10$ ) mice with tumors. \*,  $P = 0.041$ , two-tailed Fisher's exact test. D, representative colon section showing multiple tumors in the colon of an untreated mouse. Immunostaining (hematoxylin-counterstained) for MUC1 of a representative colon tumor from an untreated mouse. Scale bar, 100  $\mu\text{m}$ .



**Fig. 4.** Vaccination induces changes in the local and systemic microenvironments of MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice with IBD. **A**, representative H&E sections of colons from an untreated, an adjuvant-treated, and a vaccinated MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mouse. Scale bar, 100  $\mu$ m. Inserts are oil immersion magnification ( $\times$ 1,000) and show infiltration of neutrophils (lobular-shaped nuclei and copious cytoplasm) in the colons of untreated and adjuvant-treated mice and infiltration of lymphocytes (small round nuclei and little cytoplasm) in colons of vaccinated mice. **B**, neutrophil-produced lactoferrin levels in the feces collected at 4, 8, 10, and 16 wk of age in vaccinated ( $n = 7$ ), adjuvant-treated ( $n = 6$ ), and untreated ( $n = 5$ ) MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice, as measured by ELISA. **C**, MDSC (Gr1<sup>+</sup>CD11b<sup>+</sup>) in spleens of indicated experimental groups. Representative dot plots of several independent experiments.

10 (80%) mice that received the vaccine were tumor-free, whereas only 5 of 16 (31%) mice in the untreated group and 2 of 6 (33%) mice in the adjuvant group remained free of tumors. In Fig. 3D, we show a macroscopic view of a representative colon bearing multiple tumors from an untreated mouse. Colons in tumor-bearing mice were shortened in length, and multiple tumors were macroscopically visible. These tumors expressed high levels of MUC1, and the majority was the abnormal form (representative sections shown in Fig. 3D).

#### MUC1 vaccination changes the inflammatory microenvironment

Neutrophils are a prominent cellular component in chronic inflammation and have been shown to play a significant role in establishing a tumor-promoting microenvironment through the production of cytokines and reactive oxygen metabolites (30, 31). Representative tissue sections in Fig. 4A show a predominant granulocytic infiltrate, including neutrophils in the dysplastic lesions of untreated and adjuvant-treated mice at 16 weeks of age. In contrast, smaller mononuclear cells are the predominant population in the vaccinated mice. We postulated that the antitumor effects we observed in vaccinated mice reflected differences in the IBD cellular infiltrate and that

measuring neutrophil infiltration over time might provide a marker of that difference. We thus followed the kinetics and the extent of neutrophil infiltration using iron-binding protein lactoferrin-specific ELISA (26). Lactoferrin is stored in the secondary granules in neutrophils and secreted on their activation. Fecal samples were collected from individual mice at 4 to 5 weeks of age, early in disease and before any treatment, and then at 8, 10, and 16 weeks of age, later in the disease process for untreated mice and after treatment for the adjuvant-treated and vaccinated mice. Before treatment, mice had an average of 2,600 ng of lactoferrin per gram of feces. In all three groups, there was an increase in lactoferrin levels between 4 and 8 weeks (Fig. 4B), followed by a decrease between 8 and 10 weeks. However, between the 10-week and the 16-week time points, lactoferrin levels remained high or increased in mice that were untreated or received adjuvant alone but decreased in the vaccinated group. This decrease followed the third vaccine treatment and corresponds to the plateau in the development of rectal prolapse in the vaccinated group shown earlier in Fig. 1A.

In addition to differences in the local microenvironment, we were interested in potential systemic differences that may account for different disease outcomes between the groups. Myeloid-derived suppressor cells

(MDSC) represent another cell population that is associated with chronic inflammation and a tumor-promoting microenvironment. MDSCs are a heterogeneous population (32) characterized by the coexpression of Gr1 and CD11b on the cell surface. MDSCs have been shown to facilitate tumor development by exerting suppressor functions on an adaptive immune response (33, 34). These cells accumulate and/or expand in the secondary lymphoid organs of mice developing IBD (35). Single-cell suspensions were prepared from the spleens of vaccinated, adjuvant only, and untreated mice at 16 and 24 weeks of age. At 16 weeks of age, the untreated mice had over 3-fold higher percentages of MDSC in the spleen compared with vaccinated mice and adjuvant-treated mice (Fig. 4C). At 24 weeks, however, there was a 3.5-fold increase in spleen MDSC in adjuvant-treated mice compared with vaccinated mice and a 12-fold increase in untreated controls.

## Discussion

Using IBD and CACC as a paradigm for chronic inflammation-associated carcinogenesis, and MUC1<sup>+</sup>/IL-10<sup>-/-</sup> mice, a new mouse model that spontaneously develops these diseases, we show that inflammation can be controlled and cancer prevented by strengthening adaptive immunity against an antigen that is present at the disease site (the IBD-affected colon) and on the future malignancy (CACC). We and others have previously shown that abnormal MUC1 expression is not limited to fully transformed cells but that it can also be seen in chronic inflammation (8, 36–38). Thus, in addition to being a tumor-associated antigen, abnormal MUC1 is also an IBD-associated antigen. We also previously reported that in the well-established IL-10<sup>-/-</sup> mouse model of IBD, addition of MUC1 expression had an enhancing effect on IBD development, degree of inflammation, and progression to CACC (8). This showed that abnormal MUC1 expression was not only a marker but also an active participant in the disease that could perpetuate chronic inflammation and drive cancer development at the disease site. In the current study, we used MUC1 as the IBD-associated antigen and a MUC1 vaccine to elicit and deliver adaptive immune response (anti-MUC1) to the site of chronic inflammation (e.g., IBD), otherwise characterized primarily by the presence of innate immune effectors. By changing the balance of innate and adaptive immunity at the disease site, we expected to change the nature of ongoing inflammation and alter the course of IBD as well as its cancer-promoting potential.

We clearly show that vaccinated mice have a very different outcome than the controls in the course of their IBD, showing lesser inflammation and longer time to clinical disease. Furthermore, the controls progress to dysplasia and cancer and the vaccinated mice do not. Our data suggest that this may be the result of two different mechanisms: one a direct effector mechanism mediated by MUC1-specific adaptive immunity (anti-MUC1

IgG and MUC1-specific CTL) that seems to eliminate abnormal cells marked by abnormal expression of MUC1, and the other an indirect mechanism resulting from the changes in the local and systemic microenvironments. Locally, the protumor environment of the inflamed colon, characterized by an ever-increasing neutrophil infiltration, seems to be replaced in vaccinated mice by an environment where neutrophils decrease and mononuclear cells dominate. Similarly, the systemic protumor environment in the mouse with IBD, characterized by an ever-increasing number of MDSC in the spleen, is replaced in vaccinated mice by an environment lacking these cells.

Abnormal expression of normal cellular molecules can come about through many different events that affect normal cell functions (e.g., viral and bacterial infections, acute and chronic inflammations, and malignant transformation; ref. 39). We have previously published that some events affecting epithelial cells, such as mastitis affecting the MUC1<sup>+</sup> breast duct (37) or mumps infection of the MUC1<sup>+</sup> salivary gland,<sup>5</sup> could lead to abnormal expression of MUC1 and an immune response against it. This immunity furthermore correlated to a reduced lifetime risk of MUC1<sup>+</sup> ovarian cancer (40). The results we report here suggest that this naturally acquired anti-MUC1 immunity might also be protective of MUC1<sup>+</sup> IBD or its progression.

In view of the accumulating evidence that cancer can result from a chronic inflammatory disorder, we can consider early stages of chronic inflammation as a premalignant condition. Many molecules that are abnormally expressed in chronically inflamed tissues and the malignancies that arise from those tissues may be playing a significant role in both diseases and thus can be effectively used as targets for immunotherapy of chronic inflammation and prevention of cancer. In particular, we show that during the early stages of chronic inflammation, it might be feasible to change the maladaptive tumor-promoting microenvironment to one that is tumor inhibiting by vaccinating to induce adaptive immune responses specific for one or more disease-associated molecules.

<sup>5</sup> Unpublished data.

## Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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## References

- Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860–7.
- Balkwill F, Charles KA, Mantovani A. Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell* 2005;7:211–7.
- Sato Y, Takahashi S, Kinouchi Y, et al. IL-10 deficiency leads to somatic mutations in a model of IBD. *Carcinogenesis* 2006;27:1068–73.
- Borrello MG, Alberti L, Fischer A, et al. Induction of a proinflammatory program in normal human thymocytes by the RET/PTC1 oncogene. *Proc Natl Acad Sci U S A* 2005;102:14825–30.
- Guerra C, Schuhmacher AJ, Canamero M, et al. Chronic pancreatitis is essential for induction of pancreatic ductal adenocarcinoma by K-Ras oncogenes in adult mice. *Cancer Cell* 2007;11:291–302.
- Soucek L, Lawlor ER, Soto D, Shchors K, Swigart LB, Evan GI. Mast cells are required for angiogenesis and macroscopic expansion of Myc-induced pancreatic islet tumors. *Nat Med* 2007;13:1211–8.
- Lin WW, Karin M. A cytokine-mediated link between innate immunity, inflammation, and cancer. *J Clin Invest* 2007;117:1175–83.
- Beatty PL, Plevy SE, Sepulveda AR, Finn OJ. Cutting edge: transgenic expression of human MUC1 in IL-10<sup>-/-</sup> mice accelerates inflammatory bowel disease and progression to colon cancer. *J Immunol* 2007;179:735–9.
- Vlad AM, Kettel JC, Alajez NM, Carlos CA, Finn OJ. MUC1 immunobiology: from discovery to clinical applications. *Adv Immunol* 2004;82:249–93.
- Croce MV, Isla-Larrain M, Rabassa ME, et al. Lewis x is highly expressed in normal tissues: a comparative immunohistochemical study and literature revision. *Pathol Oncol Res* 2007;13:130–8.
- Adsay NV, Merati K, Andea A, et al. The dichotomy in the preinvasive neoplasia to invasive carcinoma sequence in the pancreas: differential expression of MUC1 and MUC2 supports the existence of two separate pathways of carcinogenesis. *Mod Pathol* 2002;15:1087–95.
- Ajioka Y, Allison LJ, Jass JR. Significance of MUC1 and MUC2 mucin expression in colorectal cancer. *J Clin Pathol* 1996;49:560–4.
- Furr AE, Ranganathan S, Finn OJ. Aberrant expression of MUC1 mucin in pediatric inflammatory bowel disease. *Pediatr Dev Pathol. Epub* 2008 Nov 24.
- Ho SB, Ewing SL, Montgomery CK, Kim YS. Altered mucin core peptide immunoreactivity in the colon polyp-carcinoma sequence. *Oncol Res* 1996;8:53–61.
- Hanisch FG, Ninkovic T. Immunology of O-glycosylated proteins: approaches to the design of a MUC1 glycopeptide-based tumor vaccine. *Curr Protein Pept Sci* 2006;7:307–15.
- Ryan SO, Vlad AM, Islam K, Garipey J, Finn OJ. Tumor-associated MUC1 glycopeptide epitopes are not subject to self-tolerance and improve responses to MUC1 peptide epitopes in MUC1 transgenic mice. *Biol Chem* 2009;390:611–8.
- Vlad AM, Muller S, Cudic M, et al. Complex carbohydrates are not removed during processing of glycoproteins by dendritic cells: processing of tumor antigen MUC1 glycopeptides for presentation to major histocompatibility complex class II-restricted T cells. *J Exp Med* 2002;196:1435–46.
- Aarnoudse CA, Garcia Vallejo JJ, Saeland E, van Kooyk Y. Recognition of tumor glycans by antigen-presenting cells. *Curr Opin Immunol* 2006;18:105–11.
- Carlos CA, Dong HF, Howard OM, Oppenheim JJ, Hanisch FG, Finn OJ. Human tumor antigen MUC1 is chemotactic for immature dendritic cells and elicits maturation but does not promote Th1 type immunity. *J Immunol* 2005;175:1628–35.
- van Kooyk Y, Rabinovich GA. Protein-glycan interactions in the control of innate and adaptive immune responses. *Nat Immunol* 2008;9:593–601.
- Rowse GJ, Tempero RM, VanLith ML, Hollingsworth MA, Gendler SJ. Tolerance and immunity to MUC1 in a human MUC1 transgenic murine model. *Cancer Res* 1998;58:315–21.
- Brokx RD, Revers L, Zhang Q, et al. Nuclear magnetic resonance-based dissection of a glycosyltransferase specificity for the mucin MUC1 tandem repeat. *Biochemistry* 2003;42:13817–25.
- Hawkins LD, Ishizaka ST, McGuinness P, et al. A novel class of endotoxin receptor agonists with simplified structure, toll-like receptor 4-dependent immunostimulatory action, and adjuvant activity. *J Pharmacol Exp Ther* 2002;300:655–6.
- Ishizaka ST, Hawkins LD. E6020: a synthetic Toll-like receptor 4 agonist as a vaccine adjuvant. *Expert Rev Vaccines* 2007;6:773–84.
- Przetak M, Chow J, Cheng H, Rose J, Hawkins LD, Ishizaka ST. Novel synthetic LPS receptor agonists boost systemic and mucosal antibody responses in mice. *Vaccine* 2003;21:961–70.
- Logsdon LK, Meccas J. A non-invasive quantitative assay to measure murine intestinal inflammation using the neutrophil marker lactoferrin. *J Immunol Methods* 2006;313:183–90.
- Schneck JP, Slansky JE, O'Herrin SM, Greten TF. Monitoring antigen-specific T cells using MHC-Ig dimers. *Curr Protoc Immunol* 2001;Chapter 17:Unit 17.2.
- Apostolopoulos V, Yuriev E, Ramsland PA, et al. A glycopeptide in complex with MHC class I uses the GalNAc residue as an anchor. *Proc Natl Acad Sci U S A* 2003;100:15029–34.
- Pisarev VM, Kinarsky L, Caffrey T, et al. T cells recognize PD(N)TR motif common in a variable number of tandem repeat and degenerate repeat sequences of MUC1. *Int Immunopharmacol* 2005;5:315–30.
- Gungor N, Godschalk RW, Pachen DM, Van Schooten FJ, Knaapen AM. Activated neutrophils inhibit nucleotide excision repair in human pulmonary epithelial cells: role of myeloperoxidase. *FASEB J* 2007;21:2359–67.
- Seril DN, Liao J, Yang GY, Yang CS. Oxidative stress and ulcerative colitis-associated carcinogenesis: studies in humans and animal models. *Carcinogenesis* 2003;24:353–62.
- Gabrilovich DI, Bronte V, Chen SH, et al. The terminology issue for myeloid-derived suppressor cells. *Cancer Res* 2007;67:425, author reply 426.
- Bunt SK, Sinha P, Clements VK, Leips J, Ostrand-Rosenberg S. Inflammation induces myeloid-derived suppressor cells that facilitate tumor progression. *J Immunol* 2006;176:284–90.
- Sica A, Bronte V. Altered macrophage differentiation and immune dysfunction in tumor development. *J Clin Invest* 2007;117:1155–66.
- Haile LA, von Wasielewski R, Gamrekelashvili J, et al. Myeloid-derived suppressor cells in inflammatory bowel disease: a new immunoregulatory pathway. *Gastroenterology* 2008;135:871–81, 881.e1–5.
- Andrianifahanana M, Moniaux N, Batra SK. Regulation of mucin expression: mechanistic aspects and implications for cancer and inflammatory diseases. *Biochim Biophys Acta* 2006;1765:189–222.
- Jerome KR, Kirk AD, Pecher G, Ferguson WW, Finn OJ. A survivor of breast cancer with immunity to MUC-1 mucin, and lactational mastitis. *Cancer Immunol Immunother* 1997;46:355–60.
- Longman RJ, Poulosom R, Corfield AP, Warren BF, Wright NA, Thomas MG. Alterations in the composition of the supramucosal defense barrier in relation to disease severity of ulcerative colitis. *J Histochem Cytochem* 2006;54:1335–48.
- Finn OJ. Immunological weapons acquired early in life win battles with cancer late in life. *J Immunol* 2008;181:1589–92.
- Cramer DW, Titus-Ernstoff L, McKolanis JR, et al. Conditions associated with antibodies against the tumor-associated antigen MUC1 and their relationship to risk for ovarian cancer. *Cancer Epidemiol Biomarkers Prev* 2005;14:1125–31.